

Research Journal of Pharmaceutical, Biological and Chemical Sciences

Correlation between ABO Blood Group Expression and The Giardiasis Infection in Human.

Fadia Abd Al-Muhsin AL-Khayat*

Department of Basic sciences, College of Dentistry, University of Baghdad , Iraq.

ABSTRACT

The current study aimed to determine the total rate of giardiasis infection in humans including infected rate with or without symptoms. Also, the infection rate according to gender was under consideration. The frequency distribution for ABO and Rh blood groups was assessed to seek for the possible relationships with giardiasis. The current study was carried in the Microbiology laboratory of Al-Yarmouk teaching hospital/Baghdad/Iraq for the period from the 1st of March till the end of August /2016. The study included 532 patients (318 male and 214 female) aged 20-45 years old to detect the existence of *Giardia lamblia* in stool samples by microscopic examination using flotation method and staining method (Lugol's iodine and Chromo trope 2R). The blood samples were collected from 130 patients for the purpose of measuring the concentration of IgG and IgM antibodies in serum using Enzyme Linked Immunosorbent Assay (ELISA) and to determine blood type using ABO and Rh blood group system. The results revealed that the total infection rate was 32.89% with no significant differences according to gender as the higher rate was recorded for males 18.79% compared with females 14.09%. For the infection rate according to state, the symptomatic patients recorded 69.7% (59.8% for males and 40.1% for females) while, asymptomatic patients recorded 30.2% (50.9% for males and 49.05% for females), no significant association $p=0.27$ between gender and the state was observed while, significant differences $P<0.0001$ for the total symptomatic and the total of asymptomatic was recorded. The mean concentration of IgM and IgG showed significant differences between infected (symptomatic, asymptomatic) and non-infected patients for each of males and females. The overall blood-group frequencies for symptomatic were A(40%) > B (26%) > O (24%) > AB (10%) and for asymptomatic were O (42%) > A (24%) > B (22%) > AB (12%) also higher percentage was observed for Rh positive 98%, 96% for symptomatic and asymptomatic respectively, no significant association between ABO $P=0.19$, Rh $P=1.04$ blood group and the state of infection was detected. A significant correlation ($P<0.01$) showed between the total infection rate and the distribution in each of blood group (0.77) and Rh test (0.79). Also a significant correlation ($P<0.01$) (0.69) and (0.59) was found between ABO and each of symptomatic and asymptomatic infections respectively. The correlations of Rh with symptomatic (0.82) and asymptomatic (0.54) were significant ($P<0.01$). The overall findings show a highly prevalent of *G. lamblia* mainly in males. There was a correlation between human blood and Rh types with giardiasis as most blood type A individuals were symptomatic patients while most blood type O were asymptomatic.

Keywords: *Giardia lamblia*, Diagnostic methods, Prevalence rate, ABO blood group and Rh

*Corresponding author

INTRODUCTION

One of the most common and important hygiene problems worldwide are parasitic infections, particularly in the developing countries[1].The World Health Organization (WHO) reports showed that people worldwide are infected with parasites, among which *G.lamblia* is the most common[2].Giardiasis is a gastrointestinal infection in humans caused by the protozoan parasite *G.lamblia*, and considered as a neglected disease in both industrialized and developing countries[3,4].The prevalence of giardiasis is as high as 10% - 50% in the developing countries [5].TheWHO has estimated more than 280 million of human infections occur annually in Africa,America and Asia, [6]. The Centers for Diseases Control (CDC) recorded that in the United States there are more than 2.5 million cases of giardiasis occur each year[7].Infected persons with Giardiasis may be showsymptoms such as diarrhea, with abdominal cramp, anorexia, and weight loss. Approximately 2×10^8 people have symptomatic giardiasis inAfrica, Latin America and Asia, and about 500,000 new cases are recorded annually[8].On the other hand, asymptomatic persons characterized by a periods with constipation or diarrhea and malabsorption[9].Epidemiology rates for asymptomatic infections according to American Medical Association [10]were about two-third of infected people have no symptoms which leads to difficulty in the eradication and control of disease [11,12] .

Because of the high incedance and disease burden of the infection, Giardiasis exerts a significance public health impact. Infections usually associated with poor personal hygiene, poor water quality and over crowding. Disease variabilityis related with multiple factors such as host age and gender, level of host immunityand nutrition, strain genotype, infective dose and possibly co-infection [13,14].

Fecal-oral rout with the ingestion of the infective stage(cyst) in contaminated food and water consider to be the main source for Giardiasis, transmission also can occur from person-to-person [15].

The detection of *G.lamblia* depended mainly on traditional method such as microscopic methods including direct smear method, Flotation method and Sedimentation method[16,17,18]. The more sensitive and more specific method focusing on the detection of Giardiaantigen using immune diagnosis through Enzyme Immunoassay(EIA) and Enzyme-Linked Immuno sorbent Assay(ELISA)[19,17].On the other hand ELISA method used for antibodies detection in serum of infected patients, the detection of specific IgG and IgM antibodies in serummay be helpfulin differentiation between patients with recent or acute infection fromthose with previous or treated infections[20].Recentlymolecular diagnosis permit the detection of causative agent depending on gene amplification such as Polymerase Chain Reaction (PCR) method [21].

Many microorganisms , parasites and virus infections associated with the differences in blood group antigen expression of the host. Blood group play a direct role indecrease orincrease host susceptibility to infections through serving as receptors and/or co-receptors and modify the natural immune response to infection.

A series of glycolipids withglycoprotein on the surface of red blood cell constitute blood group antigens ABO and Rh in rhesussystems. This antigen is genetically controlled [22]. Human populations share the same blood group systems with inherited from common ancestor but they differ by their frequencies [23].

The frequency distribution of ABO and Rh blood groups was assessed to seek for the possible relationships with giardiasis in both symptomatic and asymptomatic patients in this study.Investigation of total prevalence rate of *G.lamblia* infection in humanand the infection rate according to gender in symptomatic and asymptomatic patients was in account.

MATERIAL AND METHODS

Stool samples

During the period from the 1st of march till the end of August /2016 ,532 human stool samples were collected in sterile plastic cups approximately 5 g from patient attended Al-Yarmouk teaching hospital laboratory including 318 sample from male and 214 sample from female aged 20-45 years old, the presence of *G.lamblia* was observed in both symptomatic and asymptomatic patients.

Identification of Giardia lamblia

Macroscopic examination was done for each stool sample including the observation of the color, odour, consistency (hard, loose or watery), presence of blood, mucus and other foreign bodies.

Microscopic examination involve: direct microscopy by using saline(0.85% of NaCl solution)and 1%Lugol's iodine preparation, examination under 400X magnification to observe the trophozoite and/or cyst stage of G.lamblia. Also flotation technique which is based on specific gravity was done by using Zinc sulfate solution for cyst purification[24]. Chromotrope 2R stain(Weber's modified trichrome,1g chromotrope 2R (sigma); 0.15g fast green; 0.25g phosphotungstic acid; 3 ml acetic acid; 100 ml distilled water) was used for fecal smear staining.The development of this staining method was obtained in CDC using various components of the Trichrome staining method to differentiate G.lamblia cysts and trophozoites from background fecal elements.

Reagents of this stain include Absolute methanol, Chromotrope stain(sigma company), Acid alcohol, ethanol (95% , 100%) and Xylene. The first step of this method require fixation of fecal smear for 1-2 minutes with absolute methanol. The fecal smear then was covered for 90 minutes with Chromotrope stain then, adding acid alcohol for 1 to 3 seconds. The smear was dipped in 95% of ethanol followed by exposure to 100% of absolute ethanol for 3 minutes and transferred to xylene for 10 minutes. Slide was kept to dry in air and mounted with coverslip using Canada balsamas mounting media . Examination under microscope was done using higher power (40X) o. G.lamblia appear with light brownish to grey color [25].

Blood samples

Approving obtained from 130 patients to withdraw 3 ml of blood sample. These samples divided into three categories of 50 samples belonging to both symptomatic and asymptomatic patients (included 25 samples of both males and females) and 30 sample for non-infected persons (control) including 15 sample for both males and females. Blood was collected with a sterile, disposable plastic syringe.

ABO and Rh test

Four drops were used for blood grouping and Rh factor examination by the slide method depending on antigen-antibody agglutination reaction the blood group that showed agglutination were considered to be positive for a particular blood grouping reagent (26).

Enzyme-Linked Immunosorbent Assay (ELISA)

The rest from the blood sample was placed in Gel and Clot Activator Non Blood Collection Tube, serum was separated by centrifuge 3000 rpm for 5 minutes and stored in eppendorf tube at -20°C until use for ELISA test. According to the manufacturer's instructions the antibodies concentration of IgM and IgG in symptomatic and asymptomatic patients respectively was obtained and it was compared with the concentration found in non-infected patients serum. (Epitope Diagnostics /Catalog No: KT-846 for IgG and KT-847 for IgM)

Statistical analysis

The statistical analyses for all obtained data in the current study were undertaken using SPSS program version 21. The prevalence rate and the degree of association between blood type and the diseases were tested by using of the Chi-square test. The correlation coefficient was estimated. One way ANOVA was used in analysis of IgM and IgG and the differences among means were compared by using LSD. Probability of occurrence by chance is significant if $P < 0.05$.

RESULTS

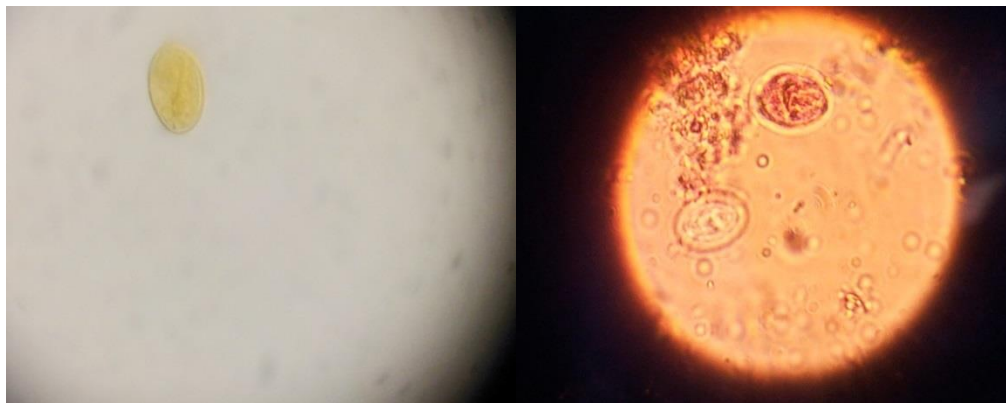
Data analysis revealed that the prevalence of G.lamblia among 532 stool samples examined under microscope was 32.89%, G.lamblia presence was observed in 100 stool samples of a total 318 examined samples for males with higher infection rate 57.14% of all positive samples and 18.79% of all total samples. On

the other hand, of a total 214 stool samples for females, G.lamblia was detected in 75 sample with 42.85%, 14.09% as infection rate of all positive and total samples respectively. Significant differences was observed according to gender.(Table 1,Figure A,B)

Table 1: Prevalence of Giardia lamblia

Gender	No. of examined samples	No. of positive	Prevalence of positive %	Prevalence of total %
male	318	100	100/175(57.14)	100/532(18.79)
female	214	75	75/175(42.85)	75/532(14.09)
total	532	175	175/175(100)	175/532(32.89)

*Chi-square =0.75 P=0.38 Significant



A-Giardia lamblia cyst stained with Lugol's Iodine B-Giardia lamblia cyst stained with chromotrop2R

In the present study most samples that diagnosed with G.lambliawere among symptomatic patients122(69.7%) including 73(59.8%) for malesand 49(40.1%) for females while, asymptomatic patients showed 53(30.2%) with 27(50.9%) for male and 26(49.05%) for females. No significant differences were detectedbetween gender and the state.

The infection rate of human giardia was detected in both symptomatic and asymptomatic patients withevidence that theasymptomatic cases represent a major proportion of the total infections rate.(Table 2)

Table 2: The prevalence of Giardia lamblia in symptomatic and asymptomatic patients

State	Male	Rate %	Female	Rate %	Total	Total Rate %
symptomatic	73	73/122(59.8)	49	49/122(40.1)	122	122/175 (69.7)
asymptomatic	27	27/53(50.9)	26	26/53(49.05)	53	53/175 (30.2)
Total	100		75		175	

*Chi-square = 1.19 p=0.27 No significant association between gender and the state

Chi-square =27.20 P<0.0001 for the total symptomatic and the total of asymptomatic

In order to determine the acute and chronic cases for giardiasis in patients, the concentration of Ig M and Ig G antibodies were obtained in serum using ELISA method. Table 3 showed the mean concentration of IgM and Ig G antibodies for 50 symptomatic and asymptomatic patients in compered with the mean concentration for 30 non infected persons. Statistical analysis revealed significant differences(p<0.05) between the mean concentration of IgM for 25 male 4.22 µ /ml , 25 female 4.41 µ /ml and the concentration detected in 15 of non infected male and female 1.80 µ /ml, 1.50 µ /mlrespectively. The same results was observed for IgG antibody measurement in which the higher mean concentration recorded in 25 male 21.46 µ /ml in compere with 15 non infected males 3.51 µ /ml. Also, the 25 infected females showed a mean concentration for IgG 21.45 µ /ml, while the mean concentration 3.68 µ /ml was recorded for 15 non infected females.

Table 3: The concentration of IgM and IgG antibodies in serum of acute and chronic infected human with Giardialamblia(mean ± standard error)

State	No.of samples	Sex	Mean of IgM concentration μ /ml and standard error
Infected with symptoms	25	Male	4.22± 0.38 A
	25	female	4.41± 1.07 A
Non infected	15	Male	1.80± 0.70 B
	15	female	1.50± 0.53 B
State	No.of samples	Sex	Mean of IgG concentration μ /ml and standard error
Infected without symptoms	25	Male	21.46± 1.01 A
	25	female	21.45± 3.9 A
Non infected	15	Male	3.51± 0.30 B
	15	female	3.68± 1.82 B

Different capital letters denoted significant differences ($p < 0.05$) between infected and non-infected individuals

Fifty blood samples (25 male, 25 female) were obtained from patients for each of symptomatic and asymptomatic for blood group test. The results indicate that for symptomatic patients higher rate 40% was associated with type A, followed by patients have type B (26%) and type O (24%) with less rate for type AB (10%) while, higher rate of type O was observed in asymptomatic patients (42%) followed by type A, B and AB 24%, 22% and 12% respectively. No significant differences were obtained between symptomatic and asymptomatic patients according to ABO and Rh test, the results showed that 98% have Rh + with only 2% have Rh- in symptomatic in compared with asymptomatic that recorded 96% for Rh+ and 4% for Rh- (Table 4)

Table 4: Blood group distribution among Symptomatic and asymptomatic patients

Blood group	Symptomatic	%	Asymptomatic	%	Total %
A	20	20/50(40)	12	12/50(24)	32
B	13	13/50(26)	11	11/50(22)	24
AB	5	5/50(10)	6	6/50(12)	11
O	12	12/50(24)	21	21/50(42)	33
total	50		50		100
Rh test					
Rh +	49	49/50(98)	47	47/50(94)	96
Rh-	1	1/50(2)	3	3/50(6)	4
Total	50		50		100

*Chi-square 4.71 $p = 0.19$ No significant association between blood group and type of infection

*Chi-square of Rh test = 1.04 $p = 0.30$ No significant association between Rh group and type of infection

Chi-square = 9.00 $P = 0.02$ for the differences among blood group in symptomatic

Chi-square = 9.30 $P = 0.02$ for the differences among blood group in asymptomatic

Chi-square = 46.08 $P < 0.0001$ for the differences between Rh group in symptomatic

Chi-square = 38.72 $P < 0.0001$ for the differences between Rh group in asymptomatic

This study is the first one in Baghdad to find if there is a relationship between the type of blood group of infected human and the state of infection (symptomatic, asymptomatic). The Pearson correlation coefficient was estimated.

Results indicated that correlation coefficient (r) recorded a significant ($P < 0.01$) between total infection rate and the distribution in each of blood group 0.772 and Rh test 0.798 respectively. The correlation between the type of blood group and infection was 0.696 for ABO, 0.822 for Rh and on the other hand, infection without symptoms showed 0.596, 0.542 for both ABO and Rh respectively.

In table 5, the distribution of blood group according to gender showed no significant differences between symptomatic and asymptomatic patients. Type A was the higher percentage 40% in the symptomatic for each of male and female as well as types A recorded 24% for both males and females in asymptomatic patients. Type B percentage was 20% for both symptomatic and asymptomatic males in compared with females that showed 32% for symptomatic and 24% for asymptomatic. The percentage of type AB in males and females for symptomatic, asymptomatic patients were 12%, 8%; 8%, 16% respectively. Asymptomatic patients showed higher percentage for type O in both males 48% and females 36% while the percentage in symptomatic males and females was 28% and 20% respectively.

Table 5: Blood group distribution in symptomatic and asymptomatic patients according to gender

Blood group	Symptomatic				Asymptomatic			
	male	%	female	%	male	%	female	%
A	10	40	10	40	6	24	6	24
B	5	20	8	32	5	20	6	24
AB	3	12	2	8	2	8	4	16
O	7	28	5	20	12	48	9	36
Total	25	100	25	100	25	100	25	100

*Chi-square=1.22 P=0.74 to measure the association between gender and blood group in symptomatic

*Chi-square=1.18 P=0.75 to measure the association between gender and blood group in asymptomatic

Chi-square=4.28 P=0.23 male symptomatic

Chi-square=5.88 P=0.11 female symptomatic

Chi-square=8.44 P=0.03 male asymptomatic

Chi-square=2.04 P=0.56 female asymptomatic

Results in table 6 revealed that Rh positive was observed in most participants, males percentage was 96% (24/25) in both symptomatic and asymptomatic while, females showed 100% for symptomatic and 92% (23/25) for the asymptomatic. On the other hand the proportion of the Rh negative individuals affected with giardiasis was 4% (1/25) in symptomatic, asymptomatic males and was 8% in asymptomatic females. Significant difference P<0.05 in the frequency of Rh factor between gender and blood group was recorded for symptomatic population.

Table 6: Rh test according to gender

Blood group	Symptomatic				Asymptomatic			
	male	%	Female	%	male	%	female	%
Rh +	24	96	25	100	24	96	23	92
Rh-	1	4	-	0	1	4	2	8
Total	25	100	25	100	25	100	25	92

*Chi-square=1.02 P=0.30 to measure the association between gender and blood group in symptomatic

*Chi-square=0.35 P=0.55 to measure the association between gender and blood group in asymptomatic

Chi-square=21.16 P<0.0001 male symptomatic

Chi-square=25.00 P<0.0001 female symptomatic

Chi-square=21.16 P<0.0001 male asymptomatic

Chi-square=17.64 P<0.0001 female asymptomatic

DISCUSSION

The higher total infection rate 32.89% for giardiasis recorded in this study was associated with many factors such as high resistance of the infective stage (cysts) to environment factors, the low infected dose (10cyst), modes of transmission (mainly inadequate clean water) and poor personal hygiene [27, 28].

Nearly closely result in the total prevalence rate was observed in Egypt, since the results of a study conducted by Foronda et al [29] revealed 34.6% as giardiasis infection rate. The CDC recorded that in developing countries the infection rate of giardiasis was 33% [30]. The difference in the results can be attributed to the number of samples examined, methods for detection, environmental differences and host immunity conditions [31].

According to patients gender, the results showed that sex of the patient had no significant ($p < 0.05$) influence on the total infectivity rate with the parasite and this has been confirmed by the results of some previous studies [32, 33]. A study done on the prevalence rate of *G. lamblia* by Amjed [34] showed males were recorded high prevalence 2.18% than females 1.51%. Another study documented the total prevalence rate of *G. lamblia* in 756 examined stool sample was 148 (19.57%) and males recorded higher rate 20.31% than female 18.11% [35].

Analysis of the results showed that males infection rate 18.79% was higher than females 14.09%. The reason may be due to the difference in the chance of exposure to the parasite between males and females through transmission modes such as food and water supplies, and other effected factors such as community hygiene, sanitation, reported household pet contact especially cats, dogs and cattle. Recently, studies focus on sex-associated hormones which may modulate immune responses and consequently affected the susceptibility to parasitic infection. [36, 37].

On the other hand, disapproval was observed with the results of several studies in that females recorded higher prevalence rate than males [38, 39, 40, 41].

According to American Medical Association, approximately two hundred million person in Asia, Africa, and Latin America have symptomatic giardiasis [42]. While about two-third of infected people have no symptoms (asymptomatic infections) which leads to difficulty in the eradication and control of disease [11, 12]. Although the proportion of infected persons without symptoms less than the proportion of persons with symptoms, asymptomatic cases of *Giardia* represent a major proportion of the total cases of infection and so giardiasis mostly occurs sporadically. In a study conducted in northern Ghana, *G. lamblia* were observed more than twice as frequently in asymptomatic individuals 12 (9.7%) than in symptomatic individuals 9 (3.7%) [43]. Also the American Academy of Pediatrics revealed that asymptomatic infection is common, approximately 50% to 75% of infected people [44]. The differences in the severity of infection due to host defenses against *Giardia* which can be classified into two categories— non-immunological responses and immunological responses [45].

There has been increasing evidence that blood groups have a possible function and biological role in disease association including bacterial, viral or parasitic infections.

The red blood cells and some human cells and tissues showed the expression of ABO antigens (A, B and H) which are complex carbohydrate molecules. So, RBCs may be used as a tool in the detection of the causative microorganisms for diseases [46, 47, 48, 49]. These carbohydrate consider to be as receptors for ligands, viruses, bacteria and parasites, enzymes, adhesion molecules and structural proteins [50]. This blood antigens may be presence or lack in some blood groups causing changes in blood cells membrane, functionally and morphologically [51].

Results of data analysis of this study showed no significant association between the ABO and giardiasis infection this may be related to the level of immunoglobulin in the serum of individual, type A was more susceptible than O, B, AB in symptomatic patients while, most of the asymptomatic patients showed O type. Only a few studies have investigated the information on the association between giardiasis and blood groups. In Egypt, a studies concluded that blood group A was more susceptible to giardiasis specially asymptomatic

form[52,53] .Al-Taiet al [54] recorded that there was no correlation between ABO and Rh blood group and giardiasis infection. Our findings were contrary to another study conducted by Ayeleetal[55].

CONCLUSION

Giardiasis infection in Baghdad still recorded a high rate in individual without any outbreak documented. Gender had a role in the incidence of infection as males were the most infected in comparison with females.

ABO may influence the risk of giardiasis by unknown mechanisms. It is now clear that ABO blood types and Rh are not the exact cause of diseases but they affect susceptibility and resistance to disease and health. Most symptomatic patients showed A blood type while, O blood type was the higher in asymptomatic patients.

RECOMMENDATIONS

More Studies on the association of *G. lamblia* genotypes with the spectrum of symptoms or asymptomatic giardiasis are required. Additional testing is needed to determine the extent of association of the blood groups genotypes with giardiasis infection. Persons with high risk blood types must be screened in order to modify their lifestyles and health behavior to prevent or control the infection.

REFERENCES

- 1 Thomas K, Fomefret Y, Emmanuel E, Therese N, Roger M and Albert S. Prevalence and Risk Factors of Fomestinal Helminth and Protozoa Infections in an Urban Setting of Cameroon: the Case of Douala. *American Journal of Epidemiology and Infectious Disease* 2015;3(2): 36-44.
- 2 World Health Organization... Working to overcome the global impact of neglected tropical diseases: First who report on neglected tropical diseases. Geneva: World Health Organization; 2010.
- 3 Eckmann L, Gillin FD. Microbes and microbial toxins: paradigms for microbial-mucosal interactions I. Pathophysiological aspects of enteric infections with the lumen-dwelling protozoan *Giardia lamblia*. *Am J Physiol Gastrointest Liver Physiol* 2001;280:G1-G6.
- 4 Daly ER, Roy SJ, Blaney DD, Manning JS, Hill VR, Xiao L and Stull JW. Outbreak of Giardiasis associated with a community drinking-water source. *Epidemiol. Infect* 2010; 138(4):491- 500.
- 5 Al-Mekhlafi HM, Al-Maktari MT, Jani R, Ahmed A, Anuar TS, Moktar N, Mahdy MA, Lim YM, Mahmud R and Surin J. Burden of *Giardia duodenalis* infection and its adverse effects on growth of school children in rural Malaysia. *PLoS Negl Trop Dis* 2013; 7(10): e 2516.
- 6 Comité OMS d'Experts. Importance des parasitoses intestinales. *Bulletin de la Organisation Mondiale de la Santé* 1988; 66(1): 23-34.
- 7 Centers for Disease Control and Prevention [CDC]. Giardiasis [online]. CDC; 2010 Nov. Available at: <http://www.cdc.gov/parasites/giardia/>. Accessed 7 Sept 2012.
- 8 Pestechian N, Rasekh H, Rostami-Nejad M, Yousofi HA and Hosseini-Safa, A. Molecular identification of *Giardia lamblia*; is there any correlation between diarrhea and genotyping in Iranian population? *Gastroenterol Hepatol Bed Bench* 2014; 7: 168-72.
- 9 Rana SV, Bhasin DK and Vinayak VK. Lactose hydrogen breathe test in *Giardia lamblia* positive patients. *Dig Dis Sci* 2005; 50:259-261.
- 10 American Public Health Association. Giardiasis (*Giardia enteritis*). In: Heymann D Ed. *Control of Communicable Diseases Manual*. 19th ed. Washington, D.C.: American Public Health Association, 2008: 258-260.
- 11 Furness BW, Beach MJ and Roberts JM. Giardiasis surveillance-United States, 1992-1997. *MMWR* 2000; 49(7): 1-13.
- 12 Davies AA, Campbell B, Evans MR, Bone A, Roche A and Chalmers RM. Asymptomatic carriage of protozoan parasites in children in day care centers in the United Kingdom. *Pediatr Infect Dis J* 2009; 28: 838- 840.
- 13 Faubert G. Immune response to *Giardia duodenalis*. *Clin. Microbiol. Rev* 2000; 13:35-54.
- 14 Alyousefi NA, Mahdy MA, Mahmud R and Lim YA. Factors associated with high Prevalence of intestinal protozoan infections among patient in Sana'a city, *Plos* 2011; 6(7): e22044.

- 15 Barry MA, Weatherhead JE, Hotez, PJ and Woc-Colburn L "Childhood parasitic infections endemic to the United States". *Pediatr Clin North Am* 2013; 60 (2): 471–85.
- 16 Stringer DA and Babyn PS. *Pediatric gastrointestinal imaging and intervention*. 2nd ed., B.C. Decker Inc Canada 2000: 50- 51.
- 17 Baker JR *Advances in parasitology*. Elsevier science and technology Books 2007 p.131.
- 18 Minvielle MC, Molina NB, Polverino D and Basualdo JA. First genotyping of *Giardia lamblia* from human and animal feces in Argentina, South America. *Mem. Inst. Oswaldo. Cruz. Rio de Janeiro* 2008; 103: 98- 103.
- 19 Carcia LS, Shimizu RY and Novak S. Commercial assay for detection of *Giardia Lamblia* and *Cryptosporidium parvum* in human fecal specimen 2003; 41: 209- 212.
- 20 Jones JE. *Giardiasis*. En: Balows A, Hausler WJ, Ohashi M, Turano A. *Laboratory diagnosis of infectious diseases* 1988; 1: 872-882
- 21 Mens P, Spieker N, Omer S, Heijnen M, Schallig H and Kager PA. Is molecular biology the best alternative for diagnosis of *Giardia* microscopy? A comparison between microscopy, antigen detection and molecular tests in rural Kenya and urban Tanzania, *Tropical Medicine and International Health* 2007; 12(2): 238-244.
- 22 Talukder SI and Das RK. Distribution of ABO and Rh Blood Groups among Blood Donors of Dinajpur District of Bangladesh *Dinajpur Med Col J* 2010; 3(2): 55-58.
- 23 Jorde L and Wooding S. Genetic variation, classification and 'race'. *Nat Genet* 2004; 36: 28-33.
- 24 Zajac AM, Johnson J and King SE. Evaluation of the importance of centrifugation as a component of zinc sulfate fecal flotation examinations. *Journal of American Animal Hospital Association* 2002; 38: 221- 224.
- 25 Moura H, David AS, Fernando B, Fernando CS, Sara W and Govinda SV. A new and improved 'quick-hot Gram-chromotrope' technique that differentially stains microsporidian spores in clinical samples, including paraffin-embedded tissue sections. *Archives of pathology & laboratory medicine* 1997; 121(8): 888-893 .
- 26 Kaya H, Gündoğdu M, Akarsu E, Kiki, İland Tekin B. The distribution of blood groups in Erzurum. *Medical Journal of Atatürk University* 199; 31: 20-22.
- 27 Hunter PR and Thompson RC. The zoonotic transmission of *Giardia* and *Cryptosporidium*. *Int J Parasitol* 2005; 35: 1181-1190.
- 28 Reiner DS, Ankarkler J, Troell K and Plam D. Synchronization of *Giardia lamblia* Identification of cell cycle stage specific genes and a differentiation restriction point. *Int. J. Parasitol* 2008; 38(3-4): 235-248.
- 29 Foronda P, Bargues M, Abreu-Acosta N, Periago M, Valero M and Mas-Coma, S. Identification of genotypes of *Giardia intestinalis* of human isolates in Egypt. *Parasitol. Res* 2008; 103: 1171-1181.
- 30 Centers For Disease Control and Prevention (CDC). *Giardiasis surveillance-United state, 2006-2008*, *MMWR* 2010; 59: 15-25.
- 31 Maria C, Edward R and Alverne P. Prevalence and associated risk factor for *Giardia Lamblia* infection among children hospitalized for diarrhea in Goiás state, Brazil, *Rev. Inst. Med. Trop* 2007; 49(3): 139- 145.
- 32 Al-Saeed AT and Issa SH. Frequency of *Giardia lamblia* among children in Dohuk, northern Iraq. *East. Mediter. Heal* 2005; 12(5): 555
- 33 Mohammad Y, Saminullah S and Azmat T. Frequency of *Giardia lamblia* infection in Children with recurrent abdominal pain. *J. Pak. Med. Asso* 2008; 58(4): 171- 174.
- 34 Amjed QI. Prevalence of *Entamoeba histolytica* and *Giardia lamblia* in children in Kadhimia hospital. Iraq. *J vet Med* 2012; 36(1): 32-36.
- 35 Sangram S P, Bhawna A, Anusha V and Chandrim S. Prevalence of Giardiasis in Patients Attending Tertiary Care Hospital in Northern India. *Int. J. Curr. Microbiol. App. Sci* 2015; 4(5): 339-344.
- 36 Morrell V. Zeroing in on how hormones affect the immune system. *Science* 1995; 269: 773–775.
- 37 Craig WR, William W and James A. Sex-Associated Hormones and Immunity to Protozoan Parasites. *Clin Microbiol Rev* 2001; 14 (3): 476-488
- 38 Al-Shammari S, Khoja T, El-Khwasky F and Gad A. Intestinal parasitic diseases in Riyadh Saudi Arabia: prevalence sociodemographic and environmental associates *Trop. Med Int Health* 2001; 6(3): 184-189.
- 39 Afkar M H (2005). Prevalence of intestinal parasites in children in Baghdad Al-Rusafa M. Sc. Thesis Faculty of Veterinary Medicine University of Baghdad.

- 40 HarithSJ.Study of some epidemiological aspects of giardiasis in north of Baghdad J Baghdad for sci 2011; 9(2): 251-252.
- 41 Shatha AW and Nada HAR. Prevalence of Blastocystishominis and GiardiaLamblia parasites in patient of four regions in east-south Baghdad;The Iraqi J Vet Med 2011; 35 (2): 74 – 84.
- 42 Thompson RC, Palmer CS and O"Handley R.The public health and clinical Significance of Giardia and Cryptosporidium in domestic animals .Vet J 2000; 177 (1): 18-25.
- 43 Klaus R, Ignatius R,Weitzel T,Seidu-Korkor A,Anyidoho L,Saad E,Djie-Maletz A,Ziniel P,Amoo-Sakyi F,Danikuu F,Danour S,Otchwemah RN,Schreier E,Bienzle U, Stark K and Mockenhaupt FP .Acute childhood diarrhoea in northern Ghana: epidemiological, clinical and microbiological characteristics BMC Infectious Diseases 2007; 7, 1471-2334
- 44 American Academy of Pediatrics. Giardia intestinalis (formerly Giardia lamblia and Giardia duodenalis) Infections. In: Pickering LK, Baker CJ, Kimberlin DW, Long SS, eds. Red Book: 2012 Report of the Committee on Infectious Disease, 29th ed. Elk Grove Village, IL: American Academy of Pediatrics; 2012: 333-335.
- 45 Gaetan F. Immune response to Giardia duodenalis. ClinMicrobiol Rev 2000; 13: 135-154.
- 46 Joann MM and John JM. Blood group associations with parasites, bacteria, and virusesTransfusionMedicineReviews2000;14(4):302-311.
- 47 Jason BH, Ashrafu I K,Regina C L ,David J,FahimaH, Abu S G F ,David A S,Edward T RFirdausi QI and Stephen B C. Blood group, immunity, and risk of infection with Vibrio cholerae in an area of endemicity. Infect.Immun 2005;73(11):7422-7427.
- 48 Jeremiah ZA,JeremiahTA andEmelikecFO.Frequencies of some human genetic markers and theirassociation with Plasmodium falciparum malaria in the Niger Delta. Nigeria J Vector Borne Dis 2010; 47: 11-16.
- 49 Kumar N, Nadimpalli M,Vardhan VR and Gopal SD. Association of ABO blood groups with Chikungunyavirus. Virol J 2010; 7: 140.
- 50 Cartron JP and Colin Y. Structural and functional diversity of blood group antigens. TransfusClinBiol 2001; 8(3):163-199
- 51 Sandler SG and Mallory D. Biological functions of blood groups in health and disease. Haematologia (Budap) 1995; 27(1):1-13.
- 52 Bouree P and BonnotG.Study of relationship of ABO and Rh blood group, and HLA antigens with parasitic diseases. J Egypt SocParasitol 1989; 19(1):67-73.
- 53 El-Ganayni GA,Attia RA and MotaweaSM.The relation between ABO blood groups, HLA typing and giardiasis in children. J Egypt SOC Parasitol1994;24(2):407-12.
- 54 Al-Taie H, Al-Mayah S and ThweniA.The association between human giardiasis and ABO and Rhesus blood groups. Iraqi JSci 1983; 4: 543-576.
- 55 Ayele M,Musin Kand Gary W.Association of Abo Blood Group and Rh Factor with Malaria and Some Gastrointestinal Infectious Disease in a Population of Adet and Merawi, Ethiopia, Global Journal of Biotechnology & Biochemistry 2014; 9 (4): 137-142.